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Synthesis, SAR and intramolecular hydrogen bonding pattern of 1,3,5-trisubstituted 4,5-dihydropyrazoles as potent cannabinoid CB₁ receptor antagonists

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ABSTRACT

The synthesis, structure–activity relationship (SAR) studies and intramolecular hydrogen bonding pattern of 1,3,5-trisubstituted 4,5-dihydropyrazoles are described. The target compounds **6–18** represent a novel class of potent and selective CB₁ receptor antagonists. Based on X-ray diffraction data, the orally active **17** is shown to elicit a different intramolecular H-bonding mode as compared to ibipinabant (**3**) and SLV330 (**4**).

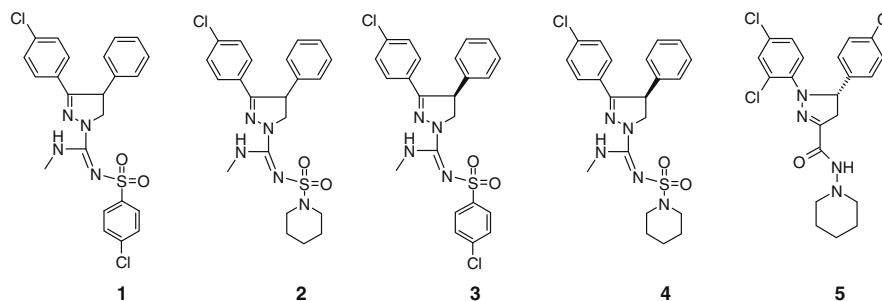
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The endocannabinoid system plays an important role in many physiological processes.¹ In particular, cannabinoid CB₁ receptor antagonists/inverse agonists have shown clinical efficacy in the treatment of obesity and related cardiovascular and metabolic risk factors,^{2,3} and have also been related^{4,5} to the potential treatment of addiction, cognitive disorders and peripherally mediated disorders like liver fibrosis, cancer, arthritis and chronic bronchitis. Although the risk of psychiatric side-effects has terminated⁶ many development programs of CB₁ receptor blockers for obesity, suggestions have been made to focus on possible therapeutic applications in peripheral pathologies⁷ and even for continuing obesity clinical trials while safeguarding the safety of patients and clinical trial subjects.⁸ The majority of the reported⁹ CB₁ receptor antagonists and inverse agonists can be described in terms of a general pharmacophore model.^{10–14}

In considering options for designing novel classes of orally active and CB₁/CB₂ subtype selective cannabinoid CB₁ receptor antagonists, the pyrazolines **1–4** were chosen as a starting point.

It was anticipated that a regioisomeric shift of the pyrazoline ring in the compounds **1–4** would lead to a novel chemotype of CB₁ receptor antagonists. As a matter of fact Solvay was the first¹⁵ to patent this principle. More recently, researchers from Esteve specifically claimed¹⁶ the optically active **5**—which contains the same *N*-piperidinylcarboxamide moiety and aryl-chloro-substitution pattern as rimonabant—in a patent application and demonstrated its *in vivo* activity in an obesity model. Its 5*S* configuration is remarkable since—based on the absolute configurations of ibipinabant¹⁷ **3** and **4**¹⁸ and their resulting threedimensional (³D) geometries—the 5*R* configuration would at forehand be expected for **5**. Srivastava et al. also embarked¹⁹ on carboxamides and hydrazides related to **5** but did not disclose the absolute configurations of their compounds. These findings prompted us to synthesize and disclose our results of target compounds²⁰ **6–18**. The activities of these novel 1,3,5-trisubstituted 4,5-dihydropyrazoles were compared with their 1,3,4-trisubstituted counterparts **1–4** and a set of novel analogues **19–25**. The focus in this study was directed to the stereochemical aspects of the resulting CB₁ receptor–ligand interactions as well as on the potential role of intramolecular H-bonding interactions in these structures. In addition, the impact of

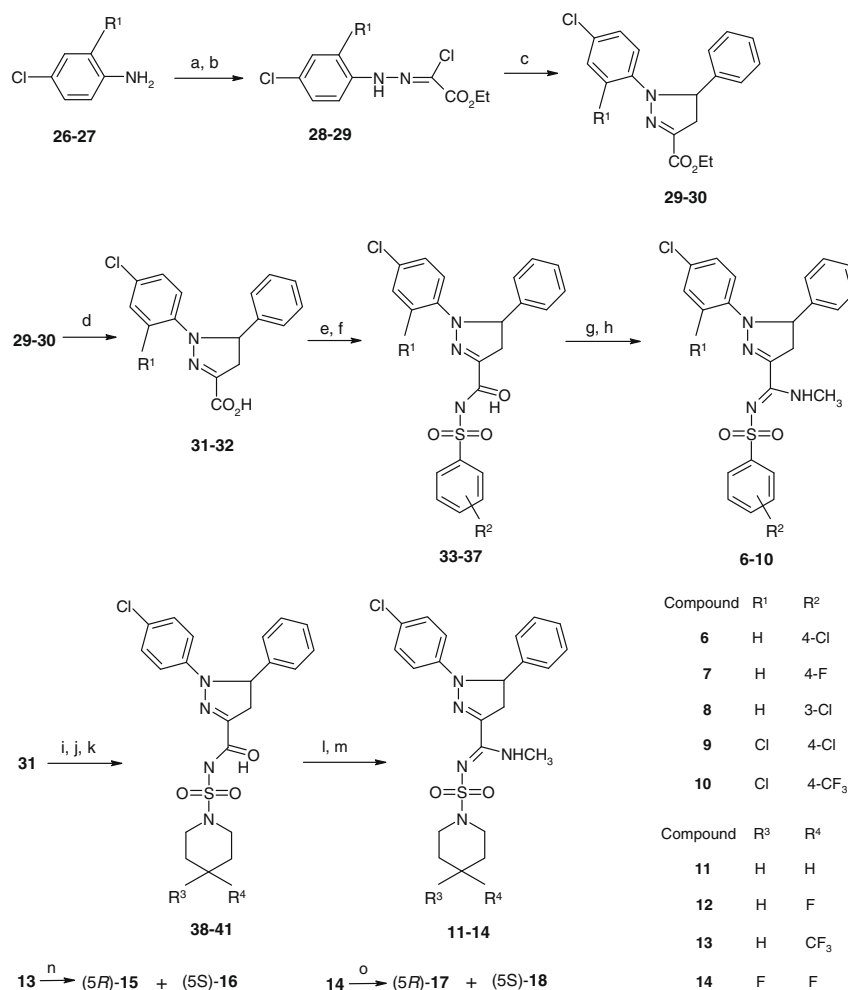
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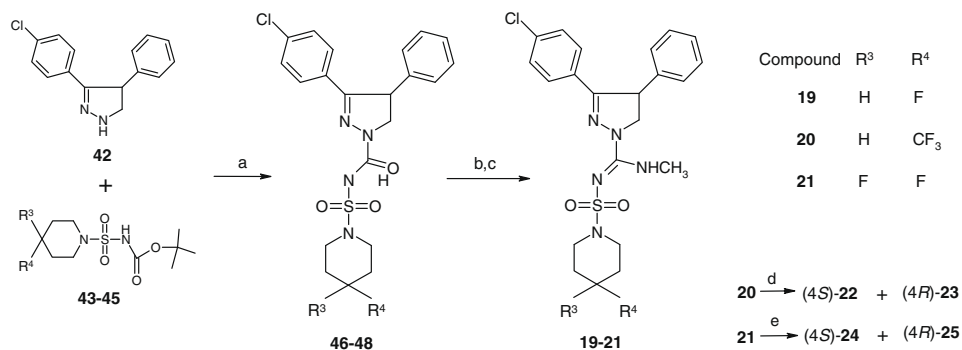
fluorination^{21–23} at the piperidine moiety of **2** was studied since this recently²⁴ led to retained CB₁ antagonism in the structurally related 1,4,5,6-tetrahydropyridazine series.

The target compounds **6–18** have been synthesized as depicted in Scheme 1. Diazotization of the anilines **26** and **27**, followed by reaction²⁵ with ethyl 2-chloro-3-oxobutanoate gave **28** and **29**, respectively. Subsequent ring closure with styrene under basic conditions at elevated temperature afforded the 3-pyrazoline esters **29** and **30** in high yields. These esters **29** and **30** were hydrolyzed under basic conditions into the carboxylic acids **31** and **32**,

which were subsequently chlorinated with SOCl₂. The in situ formed acid chlorides were reacted efficiently with aromatic sulfonamides under basic conditions to give the intermediates **33–37**. The target compounds **6–10** were obtained in varying yields by a chlorination reaction with PCl₅ at elevated temperature from **33–37**, followed by treatment with CH₃NH₂·HCl in the presence of Hünig's base. The key precursor **31** was converted analogously to crude **38–41** in quantitative yields. The conversion of the obtained crude **39–41** into the target compounds **12–14** was performed under milder chlorination conditions (POCl₃/DMAP, 40 °C) which



Scheme 1. Reagents and conditions: (a) NaNO₂, HCl, H₂O, 0–5 °C, 1 h; (b) ethyl 2-chloro-3-oxobutanoate, NaOAc, EtOH, rt, 16 h (65–73%); (c) styrene, Et₃N, benzene, reflux, 1 h (94–97%); (d) NaOH, MeOH, H₂O, reflux, 2 h (78–91%); (e) SOCl₂, toluene, 80 °C, 1 h; (f) R₂C₆H₄SO₂NH₂, CH₃CN, NaOH, rt, 16 h (78–90%); (g) PCl₅, chlorobenzene, 140 °C, 90 min; (h) CH₃NH₂·HCl, DIPEA, CH₂Cl₂, rt, 1 h (15–23%); (i) SOCl₂, toluene, 80 °C, 1 h; (j) 4,4-R³R⁴-piperidine-1-sulfamide, NaH, CH₃CN, rt, 16 h; (k) 1 N HCl, CH₂Cl₂, rt, 1 h; (l) for **11**: PCl₅, chlorobenzene, 140 °C, 90 min; for **12–14**: POCl₃, DMAP, CH₂Cl₂, reflux, 4 h; (m) CH₃NH₂·HCl, DIPEA, CH₂Cl₂, 6 °C to rt, 16 h (49–88%); (n) 250 × 30 mm CHIRALPAK[®] AD-H 5 μm column, mobile phase: 70/30 CO₂/ethanol + 1% Et₂NH (supercritical fluid chromatography), flow rate: 120 ml/min; 25 °C, detection: UV 250 nm, outlet pressure: 130 bars; (o) 250 × 76 mm CHIRALPAK[®] T-101 column, mobile phase: CH₃OH/CH₃CN/Et₂NH = 84.9/15/0.1 (v/v), flow rate: 250 ml/min; 25 °C, detection: UV 220 nm.



Scheme 2. Reagents and conditions: (a) toluene, reflux, 3 h; (b) POCl₃, DMAP, CH₂Cl₂, reflux, 4 h (70–80%); (c) CH₃NH₂·HCl, DIPEA, 6 °C to rt, 16 h (85–91%); (d) 250 × 76 mm CHIRALPAK® AD 20 μm column, mobile phase: CH₃OH/CH₃CN = 50/50 (v/v), flow rate: 270 ml/min; 25 °C, detection: UV 250 nm; (e) 250 × 80 mm CHIRALPAK® AD 20 μm column, mobile phase: CH₃OH/CH₃CN = 90/10 (v/v), flow rate: 200 ml/min; 25 °C, detection: UV 230 nm.

resulted in significantly higher yields as compared to the conversion of **38** into **11**, which was synthesized by applying the conventional reaction conditions (PCl₅, elevated temperature). To further investigate the stereochemical requirements for binding to the CB₁ receptor in this chiral pyrazoline series in more detail, the key compounds **13** and **14** were separated into their enantiomers by using chiral preparative HPLC to furnish two sets of compounds **15/16** and **17/18**, respectively.

Coupling of the dihydropyrazole building block²⁶ **42** with the carbamate esters²⁴ **43–45** afforded **46–48** in 70–80% yield, which was successively chlorinated with POCl₃ in the presence of DMAP and reacted with CH₃NH₂·HCl in the presence of Hünig's base to furnish the target compounds **19–21** in high yields (Scheme 2). The chiral compounds in this series **20** and **21** were separated into their enantiomers by using chiral preparative HPLC to furnish two sets of compounds **22/23** and **24/25**, respectively.

The pharmacological results of the compounds **1–4** and **6–25** are given in Table 1. They were evaluated in vitro at the human

CB₁ and CB₂ receptor, stably expressed into Chinese Hamster Ovary (CHO) cells,¹⁷ utilizing radioligand binding studies (displacement of the specific binding of [³H]-CP-55,940). CB₁ receptor antagonism¹⁷ was measured using a CP-55,940 induced arachidonic acid release functional assay, using the same recombinant cell line. In vivo activities of a set of key compounds after oral administration was investigated in a CB agonist-induced rat hypotension model.¹⁷

Comparison of the CB₁ receptor binding data of the novel target compound **6** and its known counterpart **1** revealed that the regioisomeric shift of the pyrazoline ring appeared to have some negative effect on the CB₁ receptor affinity of **6** (Table 1), although it should be noted that the CB₁ affinity SEM value of **6** is relatively high. However, in the case of their piperidinyl derivatives **11** and **2**, respectively, no significant impact on CB₁ receptor affinity was observed. Comparison of the CB₁ receptor affinities of the target compounds **6** and **9** showed a positive effect of the additional 2-Cl atom in **9**. In this arylsulfonyl series **6–10**, the CF₃-substituted **10** showed the highest CB₁ receptor affinity (11.3 nM). However,

Table 1
Pharmacological in vitro and in vivo results of compounds **1–4** and **6–25**

Compound	K _i (CB ₁) ^a , nM	pA ₂ (CB ₁) ^b	K _i (CB ₂) ^c , nM	CB ₁ /CB ₂ ratio	Hypotension rat ^d , ID ₅₀
1	25.2 ± 7.4	8.7 ± 0.3	>1000	>39	15
2	152 ± 68	8.7 ± 0.3	1321 ± 264	8.6	n.d. ^e
3	7.8 ± 1.4	9.9 ± 0.6	7943 ± 126	1018	5.5
4	58 ± 19	8.7 ± 0.2	3495 ± 968	60	8
6	84 ± 56	8.4 ± 0.3	>1000	>11	>30
7	21.7 ± 10.0	8.0 ± 0.3	>1000	>46	30
8	53 ± 10	8.3 ± 0.2	330 ± 171	6	>30
9	37 ± 23	9.0 ± 0.1	248 ± 96	7	>30
10	11.3 ± 2.0	8.6 ± 0.1	179 ± 38	16	>30
11	141 ± 58	7.9 ± 0.2	1086 ± 86	8	>30
12	55 ± 16	8.4 ± 0.2	>1000	>18	>30
13	7.6 ± 2.0	9.4 ± 0.3	606 ± 204	84	n.d. ^e
14	38.8 ± 14.0	9.1 ± 0.6	>1000	>25	9
15	2.0 ± 1.0	9.5 ± 0.1	584 ± 220	292	n.d. ^e
16	58 ± 22	8.2 ± 0.1	>1000	>17	n.d. ^e
17	9.2 ± 2.0	8.8 ± 0.1	>1000	>108	5
18	280 ± 81	<6.7	>1000	>3.5	>30
19	7.5 ± 4.3	9.0 ± 0.2	>1000	>133	>30
20	14.5 ± 11.9	9.4 ± 0.1	>1000	>68	n.d. ^e
21	12.2 ± 5.0	9.2 ± 0.3	219 ± 91	18	23
22	4.7 ± 1.7	9.6 ± 0.1	305 ± 58	65	2
23	140 ± 60	7.3 ± 0.2	>1000	>7	>30
24	42 ± 24	9.0 ± 0.1	>1000	>23	6
25	463 ± 168	6.4 ± 0.2	>1000	>2	>30

^a Displacement of specific CP-55,940 binding in CHO cells stably transfected with human CB₁ receptor, expressed as K_i ± SEM (nM).

^b [³H]-Arachidonic acid release in CHO cells expressed as pA₂ ± SEM values.

^c Displacement of specific CP-55,940 binding in CHO cells stably transfected with human CB₂ receptor, expressed as K_i ± SEM (nM). The values represent the mean result based on at least three independent experiments.

^d Antagonism of CB agonist (CP-55,940) induced hypotension (rat), expressed as ID₅₀ (mg/kg, po administration).

^e n.d. = not determined.

the presence in **10** of the additional 2-Cl substituent in combination with the CF₃ group led also to a highly lipophilic²⁷ compound (A log P^{28} = 5.9). Compound **7** elicited the highest CB₁/CB₂ receptor selectivity in this set of compounds.

Significant differences exist in the SAR between the 1,3,4-trisubstituted pyrazoline class and the 1,3,5-trisubstituted pyrazoline class. For example, in the novel 1,3,5-trisubstituted pyrazoline series, the 4-F substituted **7** showed a fourfold higher CB₁ receptor affinity than its 4-Cl analogue **6**. In contrast, in the earlier reported¹⁷ 1,3,4-trisubstituted pyrazoline series the 4-Cl substituted **1** showed a 13-fold higher CB₁ receptor affinity than its 4-F substituted counterpart (cf. compounds **67** and **72**, respectively, in Ref. 17). Such a marked difference in SAR cannot be rationalized based on the reported^{10–14} CB₁ inverse agonist pharmacophore model but is an indicator for distinct structural differences between both classes of pyrazolines. One of the aims of this investigation was the identification of relevant factors involved herein.

In the piperidinylsulfonyl-derived series **11–14**, the CF₃-substituted **13** elicited the highest CB₁ receptor affinity and functional CB₁ antagonistic potency. Comparison of the regioisomeric pairs **12/19** and **14/21** showed that the CB₁ receptor affinities are higher in the 1,3,4-trisubstituted pyrazoline series. However, the opposite was observed for **13** and **20**, respectively. In particular, the compounds **13** and **19** share high CB₁ receptor affinity values with high CB₁/CB₂ subtype selectivities and strong CB₁ antagonistic potencies.

Since **13** and **14** are racemates their enantiomerically pure constituents were also tested. In both cases the dextrorotatory enantiomers **15** and **17** had significantly higher CB₁ receptor affinities and CB₁ antagonistic potencies than their levorotatory counterparts **16** and **18**. The highest CB₁ receptor affinity (2.6 nM) was found in the eutomer **15**. The distomer **16** showed ~30-fold less CB₁ affinity than the eutomer **15**, indicating that these chiral ligands bind stereoselectively to the CB₁ receptor. A comparable degree of stereoselective binding to the CB₁ receptor was observed for the 1,3,4-pyrazoline counterparts **22** and **23**. The distomer **18** showed ~30-fold less CB₁ affinity than **17**, but in this case the corresponding 1,3,4-pyrazoline counterparts **25** and **24** elicited not more than an 11-fold difference in CB₁ receptor affinity.

Interestingly, the impact of variation in stereochemistry on the observed CB₁ receptor affinities in our sulfonylamidine-based series of 1,3,5-trisubstituted 4,5-dihydropyrazoles appears to be considerably higher (~30-fold ratio for the compound sets **15/16** and **17/18**, respectively) as compared with the hydrazide-based compounds,¹⁹ such as the reported analogue of **5** wherein its piperidinyl moiety was substituted by a 4-morpholinyl group (~threefold difference).

The *in vivo* activity after oral administration was investigated in a CB₁ agonist (CP-55,940) induced hypotension rat model.¹⁷ Disappointingly, our initially prepared set of arylsulfonyl-based target compounds **6–10** were hardly active or inactive in this model, whereas compound **1** previously had shown significant activity therein. Therefore, we focused our attention on the piperidine-based compound **11**, but this compound was also found inactive after oral administration. Based on molecular modeling studies it became apparent that the CB₁ receptor would accommodate an additional substituent at the 4-position of the piperidine ring of **11**. This prompted the design of the compounds **12–14**. Gratifyingly, the 4,4-difluoro substituted **14** in this series was found orally active (ID₅₀ = 9 mg/kg). According to expectation, its related eutomer **17** was approximately twofold more active in this model after oral administration (ID₅₀ = 5 mg/kg). In our piperidinylsulfonyl-based 1,3,4-trisubstituted pyrazoline series we also identified some novel compounds with a high oral activity. The 4,4-difluoro substituted **24** (ID₅₀ = 6 mg/kg) and in particular the 4-CF₃ substituted **22** (ID₅₀ = 1.9 mg/kg) were found very active in our mecha-

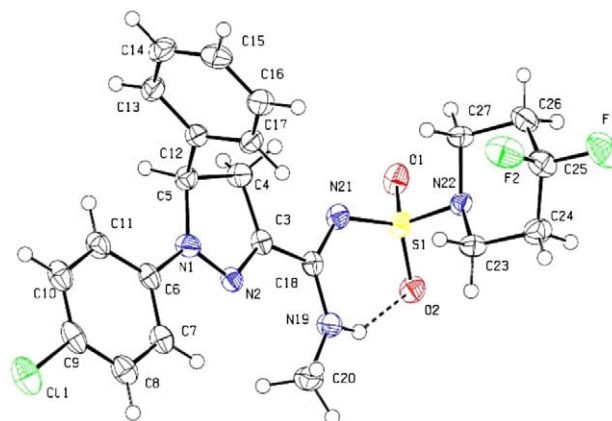


Figure 1. ORTEP drawing of the X-ray diffraction result of **17**. Its intramolecular H-bond is indicated as a dashed line.

nistic *in vivo* rodent model after oral administration. It is interesting to note that **22** was found more potent in this model than ibipinabant (**3**), as well as rimonabant¹⁷ (ID₅₀ = 3.2 mg/kg).

The effects of intramolecular H-bonding on pharmacodynamic and pharmacokinetic properties of drugs are a known phenomenon.^{29–31} Intramolecular H-bonding in general decreases the polar surface area, thereby enhancing the membrane permeability and consequently improving oral absorption. It also generally results in a somewhat higher degree of conformational constraint as compared to structural analogues lacking such an intramolecular binding feature. The comparison of the X-ray diffraction structure of **17** with the earlier reported X-ray diffraction results of compounds **3**¹⁷ and **4**¹⁸, respectively, showed that intramolecular H-bonding is present in both structure types. Both the Flack x parameter³² value (-0.06 ± 0.04) and the independent analysis of the data in terms of the Hoofstede parameter³³ (-0.028 ± 0.016) indicated an enantiomerically pure crystal of **17** with a high level of confidence. Unexpectedly, a clear difference in intramolecular H-bonding exists between the two sets of pyrazoline chemotypes. Apparently, the different sprouting of the three substituents on the common pyrazoline scaffold in **3** and **4** on the one hand and **17** on the other hand has a large impact on their preferred intramolecular H-bonding modes. An intramolecular H-bond interaction between the pyrazoline N₂ atom and its amidine N–H hydrogen atom was present in both **3** and **4**, whereas a strong intramolecular H-bond interaction between one of the oxygen atoms of the sulfonyl group and its amidine N–H hydrogen atom was observed in **17** (Fig. 1). The difference in intramolecular H-bond formation between the two sets of compounds can be rationalized by invoking the different inter-atom distance between the pyrazoline C₃ atom and the amidine C atom in **17** (1.46 Å) as compared to the pyrazoline N₁ atom and the amidine C atom in **3** (1.36 Å), respectively. The shorter distance in **3** can be explained by a higher degree of conjugation between its carboxamide moiety and the pyrazoline ring, thereby resulting in partial double bond character and a shorter bond length. These shorter bond lengths in **3** and **4**, as well as differences in their overall molecular geometry—due to their differently substituted pyrazoline ring—as compared to **17**, apparently facilitate the specific intramolecular H-bond as shown in Figure 2. The observed differences in SAR described hereinabove between some corresponding members of the two pyrazoline classes can be rationalized by invoking differences in the resulting 3D shapes, as a result of different modes of intramolecular H-bonding. More specific, an analogous orientation of both pyrazoline-bound aromatic moieties and one of the sulfonyl-oxygen atoms (which is supposed¹⁰ to form a key interaction with the Lys192 residue of the CB₁ receptor via H-bonding) at the CB₁ receptor will lead to a different positioning

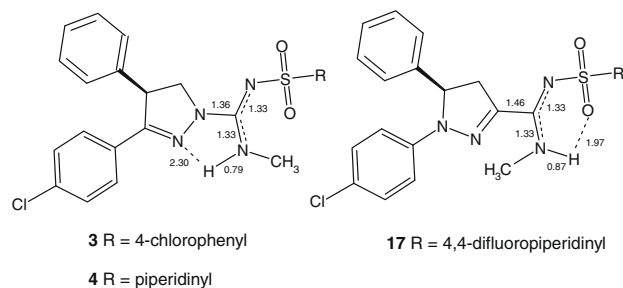


Figure 2. Schematic representation of the intramolecular H-bonding pattern in the compounds **3** and **4** versus **17**. The depicted inter-atom distances for **3** and **17** from their X-ray diffraction analyses are expressed in Å.

of both their amidine N-CH₃ substituent and the hydrophobic piperidine region therein. It should be noted that the presence of an alternative hydrophobic pocket in the CB₁ receptor has been suggested earlier.³⁴

Surprisingly, the key compound **17** showed the opposite absolute configuration as compared to Esteve's **5**. Both compounds would be expected to bind analogously to the CB₁ receptor, based on the published^{10–14} CB₁ inverse agonist pharmacophore model, since their pyrazoline moieties are almost identical and an H-acceptor moiety is present in their lipophilic tail region. It should be borne in mind that both the size and nature of their tails (N-piperidinylcarboxamide in **5** vs 4,4-difluoropiperidinyl-sulfonylcarboxamide in **17**) is different which can lead to different preferential binding modes¹⁴ at the CB₁ receptor. Additional sophisticated molecular modeling investigations will be mandatory to provide a clear explanation for these observations.

Novel 1,3,5-trisubstituted 4,5-dihydropyrazoles are described. The target compounds³⁵ **6–18** represent a novel class of potent and selective CB₁ receptor antagonists. Although the key compound **17** showed the opposite absolute configuration—according to the Cahn-Ingold-Prelog rules—as compared to **3** and **4**, the stereochemical orientation of the relevant side chains for CB₁ receptor binding in **17** matched the orientation of the corresponding substituents in **3** and **4**. The orally active **17** was shown to elicit a different intramolecular H-bonding pattern as compared to **3** and **4**. The observed clear differences in SAR described hereinabove between some corresponding members of the two pyrazoline classes was rationalized by invoking differences in ³D shape, as a result of different modes of intramolecular H-bonding as well as the different attachment points of the three substituent at their pyrazoline ring.

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- Yields refer to isolated pure products unless otherwise noted and were not maximized. Selected data for compounds **14**, **15**, **17**, **22** and **24**. *Synthesis of compound 14*: To a stirred solution of **26** (15.68 g, 0.123 mol) in ice (30 ml) and concentrated HCl (30 ml) was slowly added a solution of NaNO₂ (9.0 g, 0.13 mol) in H₂O (16 ml) and the resulting solution was stirred for 1 h at 0–5 °C and subsequently added to a cold mixture of NaOAc (32 gram, 0.39 mol), EtOH (520 ml) and ethyl 2-chloro-3-oxobutanoate (16.6 ml, 0.12 mol). After stirring the resulting mixture for 1 h the formed precipitate was collected by filtration, washed with EtOH and dried in vacuo to give **28** (22.99 g, 73% yield). Mp 147.5–149.5 °C. ¹H NMR (200 MHz, CDCl₃) δ 1.40 (t, J = 7 Hz, 3H), 4.39 (q, J = 7 Hz, 2H), 7.16 (br d, J = 8 Hz, 2H), 7.30 (br d, J = 8 Hz, 2H), 8.31 (br s, 1H). To a stirred boiling solution of **28** (22.95 g, 0.088 mol) and styrene (30.3 ml, 0.264 mol) in benzene (140 ml) was added Et₃N (34.3 ml, 0.247 mol) and the resulting solution was heated at reflux temperature for 1 h. The resulting solution was cooled to rt and the formed precipitate was removed by filtration and washed with toluene. The filtrate was concentrated in vacuo and purified by flash chromatography (silica gel, CH₂Cl₂) to give **29** (27.2 g, 94% yield) as a syrup, which slowly solidified on standing. ¹H NMR (200 MHz, CDCl₃) δ 1.38 (t, J = 7 Hz, 3H), 3.06 (dd, J = 18 and 7 Hz, 1H), 3.73 (dd, J = 18 and 13 Hz, 1H), 4.33 (q, J = 7 Hz, 2H), 5.38 (dd, J = 13 and 7 Hz, 1H), 7.02 (br d, J = 8 Hz, 2H), 7.08–7.40 (m, 7H). To a stirred suspension of **29** (23.0 g, 0.07 mol) in CH₃OH (200 ml) was added H₂O (15 ml) and concentrated NaOH (10 ml) and the resulting solution was heated at reflux temperature for 2 h. The CH₃OH was partly removed by evaporation and the residue was dissolved in a mixture of H₂O and EtOAc. Ice, concd HCl (20 ml) and EtOAc were successively added, the EtOAc layer collected, dried over MgSO₄, filtered and concentrated in vacuo. The resulting residue was washed with Et₂O (100 ml) and diisopropyl ether, respectively, to give **31** as a solid, mp 177–179 °C. To **31** (18.77 g, 62.4 mmol) in toluene (200 ml) was added SOCl₂ (18.0 ml, 246.8 mmol). The reaction mixture was stirred at 80 °C for 1 h. Volatiles were thoroughly removed in vacuo. The residue was dissolved in CH₃CN (250 ml); solution A. To a solution of 4,4-difluoropiperidine-1-sulfonamide (12.5 g, 62.4 mmol) in CH₃CN (500 ml) was added aqueous NaOH (8.25 ml, 157.8 mmol). After 10 min, solution A was slowly added. The reaction mixture was stirred overnight at rt. Volatiles were removed in vacuo to give crude **41** (39.01 g). This crude residue was extracted with CH₂Cl₂/1 N HCl. Layers were separated. The CH₂Cl₂ layer was dried over Na₂SO₄, filtered and evaporated to give **41** (30.41 g, quantitative yield). ¹H NMR (400 MHz, DMSO-d₆) δ 1.93–2.10 (m, 4H), 2.75 (dd, J = 18 and 6 Hz, 1H), 3.12–3.21 (m, 4H), 3.36 (br s, probably NH and H₂O), 3.62 (dd, J = 18 and 13 Hz, 1H), 5.42 (dd, J = 13 and 6 Hz, 1H), 6.93 (br d, J = 8, 2H), 7.14–7.36 (m, 7H). *Compound 41* (30.14 g, 62.4 mmol) was dissolved in CH₂Cl₂ (500 ml). DMAP (33.80 g, 276.7 mmol) was added. POCl₃ (7.35 ml, 80.3 mmol) in CH₂Cl₂ (50 ml) was added dropwise. The reaction mixture was heated at reflux temperature for 4 h. After cooling down to 6 °C CH₃NH₂·HCl (19.0 g, 281.4 mmol) was added, followed by dropwise addition of DIPEA (72.0 ml, 420.6 mmol). The reaction mixture was stirred overnight at rt. Water (100 ml)

was added, followed by acidification with 1 N HCl. Layers were separated. The CH₂Cl₂ layer was dried over Na₂SO₄, filtered and concentrated in vacuo. Purification by flash chromatography (silica gel, Et₂O/pa (40/60) = 1/1 (v/v)) afforded **14** (27.1 g; 88% yield.) ¹H NMR (400 MHz, DMSO-*d*₆) δ 2.00–2.15 (m, 4H); 2.85 (br d, *J* ~5 Hz, 3H), 3.09–3.21 (m, 5H), 3.94 (dd, *J* = 18 and 13 Hz, 1H), 5.61 (dd, *J* = 13 and 6 Hz, 1H), 7.04 (br d, *J* = 8, 2H), 7.20–7.38 (m, 7H), 8.80–8.85 m, 1H); ESI⁺-MS exact mass calcd for C₂₂H₂₅ClF₂N₅O₂S *m/z*, 496.1386 ([MH⁺]), found: 496.1411. **Compound 15**: ¹H NMR (400 MHz, DMSO-*d*₆) δ 1.43–1.60 (m, 2H), 1.83–1.92 (m, 2H), 2.39–2.48 (m, 1H), 2.57–2.69 (m, 2H), 2.88 (br s, 3H), 3.14 (dd, *J* = 18 and 7 Hz, 1H), 3.60 (t, *J* = 9 Hz, 2H), 3.94 (dd, *J* = 18 and 13 Hz, 1H), 5.60 (dd, *J* = 13 and 7 Hz, 1H), 7.03 (d, *J* = 9 Hz, 2H), 7.23 (d, *J* = 9 Hz, 2H), 7.26–7.39 (m, 5H), 8.86 (br s, 1H); ESI⁺-MS exact mass calcd for C₂₃H₂₆ClF₃N₅O₂S *m/z*, 528.1448 ([MH⁺]), found: 528.1475; [α]_D²⁵ = 167 (c 1 g/100 ml, CH₃OH). **Compound 17**: mp 158 °C (EtOH). ¹H NMR (400 MHz, DMSO-*d*₆) δ 2.00–2.15 (m, 4H), 2.85 (br d, *J* ~5 Hz, 3H), 3.09–3.21 (m, 5H), 3.94 (dd, *J* = 18 and 13 Hz, 1H), 5.61 (dd, *J* = 13 and 6 Hz, 1H), 7.04 (br d, *J* = 8 Hz, 2H), 7.20–7.38 (m, 7H), 8.80–8.85 m, 1H); ESI⁺-MS exact mass calcd for C₂₂H₂₅ClF₂N₅O₂S *m/z*, 496.1386 ([MH⁺]), found: 496.1403; [α]_D²⁵ = 165 (c 1 g/100 ml, CH₃OH). Selected crystallographic data for **17**: X-ray data were collected

under the supervision of Professor Dr. A. L. Spek (Bijvoet Centre for Biomolecular Research, Utrecht University, The Netherlands) with a Nonius KappaCCD diffractometer on a rotating anode using MoKα radiation, temperature: 150 K, crystal size: 0.21 × 0.30 × 0.66 mm, crystal system: monoclinic, space group: *P*21, unit cell dimensions: *a* = 6.0743; *b* = 10.4341; *c* = 17.6350 Å, Calculated density: 1.475 g cm⁻³. The CIF has been deposited at the Cambridge Crystallographic Data Centre (CCDC), deposition number: 760300. **Compound 22**: mp 164.5–165 °C. ¹H NMR (400 MHz, CDCl₃) δ 1.67–1.80 (m, 2H), 1.88–1.95 (m, 2H), 1.99–2.13 (m, 1H), 2.51–2.63 (m, 2H), 3.25 (d, *J* = 7 Hz, 3H), 3.77–3.86 (m, 2H), 4.10–4.17 (m, 1H), 4.57 (t, *J* = 11 Hz, 1H), 4.67 (dd, *J* = 11 and 5.5 Hz, 1H), 6.81 (br s, 1H), 7.15 (d, *J* = 8 Hz, 2H), 7.23–7.36 (m, 5H), 7.52 (br d, *J* = 8 Hz, 2H); ESI⁺-MS exact mass calcd for C₂₃H₂₆ClF₃N₅O₂S *m/z*, 528.1448 ([MH⁺]), found: 528.1472; [α]_D²⁵ = -130 (c 1 g/100 ml, CH₃OH). **Compound 24**: mp 185.5–186 °C. ¹H NMR (400 MHz, CDCl₃) δ 2.03–2.16 (m, 4H), 3.24 (d, *J* = 7 Hz, 3 H), 3.26–3.34 (m, 4H), 4.14 (dd, *J* = 12 and 5.5 Hz, 1H), 4.57 (t, *J* = 12 Hz, 1H), 4.67 (dd, *J* = 12 and 5.5 Hz, 1H), 6.79 (br s, 1H), 7.15 (d, *J* = 8 Hz, 2H), 7.23–7.35 (m, 5H), 7.53 (d, *J* = 8 Hz, 2H); ESI⁺-MS exact mass calcd for C₂₂H₂₅ClF₂N₅O₂S *m/z*, 496.1386 ([MH⁺]), found: 496.1399; [α]_D²⁵ = -148 (c 1 g/100 ml, CH₃OH).